



# Statistical Rigor in Genomics Data Science

in honor of Peter Bickel's 82nd birthday

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## Volume 447 Issue 7146, 14 June 2007



[Published: 14 June 2007](#)

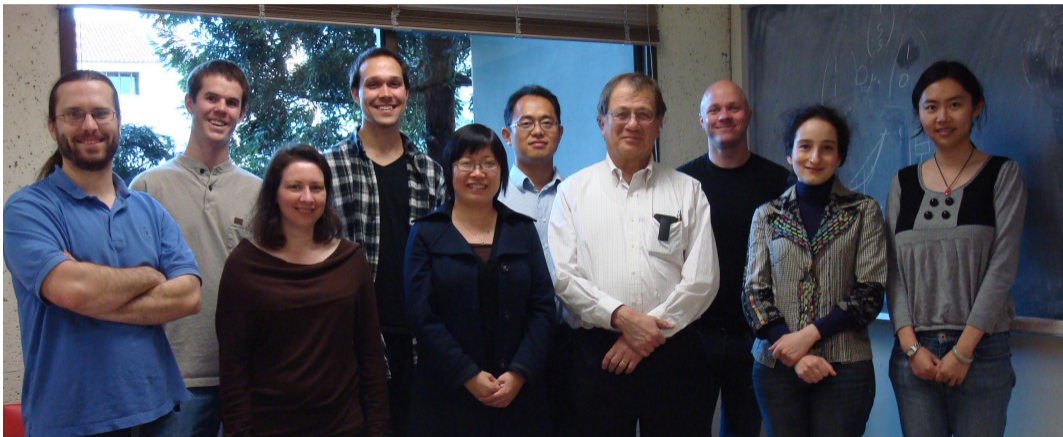
### Identification and analysis of functional elements in 1% of the human genome by the ENCODE pilot project

[The ENCODE Project Consortium](#)

[Nature](#) 447, 799–816 (2007) | [Cite this article](#)

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## Peter and Haiyan's ENCODE team (2007–2013)



# Berkeley statisticians help find function of “junk” DNA in human genome

By [Robert Sanders](#), Media relations | SEPTEMBER 6, 2012



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UC Berkeley statisticians played a key role in the large ENCODE consortium that determined the function of what was thought to be “junk” DNA in the human genome. The consortium’s 440+ scientists reported their findings in 30 journal papers on Sept. 6.

Peter Bickel, professor of statistics, was the unofficial lead statistician for the group, which involved scientists from around the world. Bickel and his UC Berkeley colleagues provided several of the tools biologists needed to uncover the functional roles of DNA outside protein coding genes.



# Our ENCODE fruits (2012–2014)

[Open Access](#) | [Published: 05 September 2012](#)

## An integrated encyclopedia of DNA elements in the human genome

[The ENCODE Project Consortium](#)

[Nature](#) **489**, 57–74 (2012) | [Cite this article](#)

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## Comparison of *D. melanogaster* and *C. elegans* developmental stages, tissues, and cells by modENCODE RNA-seq data

Jingyi Jessica Li<sup>1,3</sup>, Haiyan Huang<sup>1,4</sup>, Peter J. Bickel<sup>1,4</sup> and Steven E. Brenner<sup>2,4</sup>

[Open Access](#) | [Published: 27 August 2014](#)

## Comparative analysis of regulatory information and circuits across distant species

[Alan P. Boyle](#), [Carlos L. Araya](#), ... [Michael Snyder](#) [+ Show authors](#)

[Nature](#) **512**, 453–456 (2014) | [Cite this article](#)

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[Open Access](#) | [Published: 27 August 2014](#)

## Comparative analysis of the transcriptome across distant species

[Mark B. Gerstein](#) , [Joel Rozowsky](#), ... [Robert Waterston](#) [+ Show authors](#)

[Nature](#) **512**, 445–448 (2014) | [Cite this article](#)

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# Some questions I had about bioinformatics methods

1. Are p-values valid?
2. Why not classical statistical methods?
3. What is the proper null hypothesis?



## Criteria need calibration

- **p-values**  $\sim$  (super-)uniform $[0, 1]$  under the null hypotheses
- **false discovery rate (FDR)** =  $\mathbb{E} \left[ \frac{\# \text{ false discoveries}}{\# \text{ discoveries} \vee 1} \right] \leq$  the claimed level (e.g., 5%)



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## Three common causes of ill-posed p-values

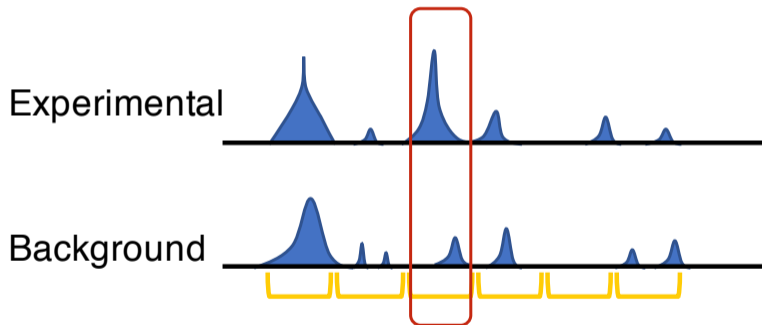
1. Formulation of a **two-sample test** as a one-sample test
2. Specification of a **parametric model** that does not fit data well
3. Treatment of **inferred** covariates as observed





# 1. Formulation of a two-sample test as a one-sample test

Example: peak calling from ChIP-seq data



# 1. Formulation of a two-sample test as a one-sample test

## Peak calling from ChIP-seq data

- Popular software:
  - MACS [Zhang *et al.*, *Genome Biol*, 2008]; cited > 10K times
  - HOMER [Heinz *et al.*, *Mol Cell*, 2010]; cited ~ 8K times

- Formulation:

a region	background count	experimental count
random variable (hypothetical)	$X$	$Y$
random observation (data)	$x$	$y$

p-value =  $\mathbb{P}(Y \geq y)$ , where  $Y \sim \text{Poisson}(x)$  — correct?



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## Peak calling from ChIP-seq data

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p-value =  $\mathbb{P}(Y \geq y)$ , where  $Y \sim \text{Poisson}(x)$  — correct?

- No, because it assumes  $Y \sim \text{Poisson}(\lambda)$  and tests

$$H_0 : \lambda = x \quad \text{vs.} \quad H_1 : \lambda > x,$$

which treats  $x$  as a fixed parameter and ignores its randomness



# 1. Formulation of a two-sample test as a one-sample test

How to perform a two-sample test when the sample size is **1 vs. 1**?

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How to perform a two-sample test when the sample size is **1 vs. 1**?

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- but, p-values are just **intermediates for FDR control** in large-scale multiple testing

**Our solution:** inspired by knockoffs [Barber and Candès, *Ann Stat*, 2015]  
(to be elaborated)

Method | [Open Access](#) | [Published: 11 October 2021](#)

## Clipper: $p$ -value-free FDR control on high-throughput data from two conditions

[Xinzhou Ge](#), [Yiling Elaine Chen](#), [Dongyuan Song](#), [MeiLu McDermott](#), [Kyla Woysner](#), [Antigoni Manousopoulou](#), [Ning Wang](#), [Wei Li](#), [Leo D. Wang](#) & [Jingyi Jessica Li](#) 

[Genome Biology](#) **22**, Article number: 288 (2021) | [Cite this article](#)

**6169** Accesses | **4** Citations | **52** Altmetric | [Metrics](#)

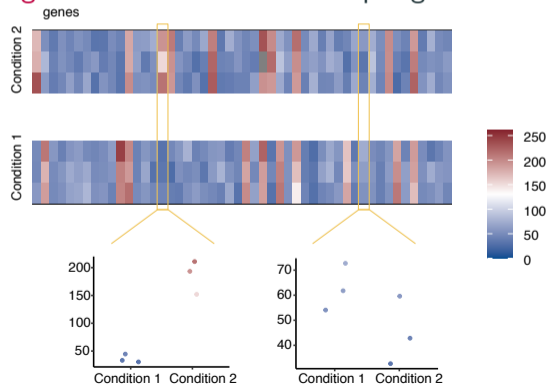


## 2. Specification of a parametric model that does not fit data well

### Example: identifying differentially expressed genes (DEGs) from RNA-seq data

- Popular software (originally designed for **small** sample sizes):
  - edgeR [Robinson *et al.*, *Bioinformatics*, 2014]; cited  $\sim 24\text{K}$  times
  - DESeq2 [Love *et al.*, *Genome Biol*, 2014]; cited  $> 33\text{K}$  times

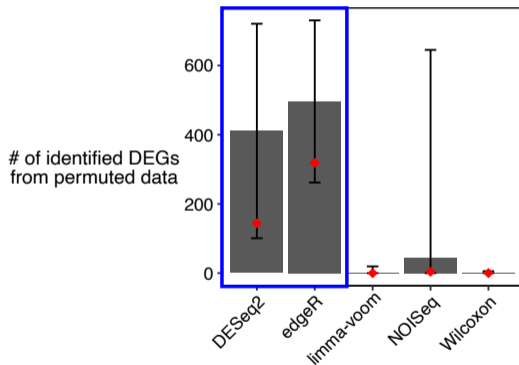
both assume a **negative binomial** distribution per gene and condition



## 2. Specification of a parametric model that does not fit data well

### Identifying differentially expressed genes (DEGs) from RNA-seq data

- Check of false discoveries: permute individuals between conditions (no true DEGs)



◆ # of identified DEGs from the original data

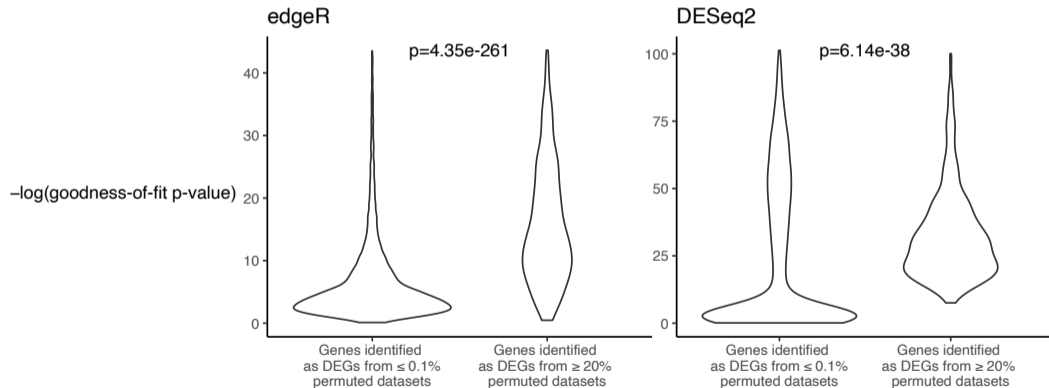
51 pre-nivolumab and 58 on-nivolumab anti-PD-1 therapy patients [Li et al., *Genome Biology*, 2022]  
[Riaz et al., *Cell*, 2017]



## 2. Specification of a parametric model that does not fit data well

### Identifying differentially expressed genes (DEGs) from RNA-seq data

- Poor fit of **negative binomial model**  $\longleftrightarrow$  false positive DEGs



51 pre-nivolumab and 58 on-nivolumab anti-PD-1 therapy patients

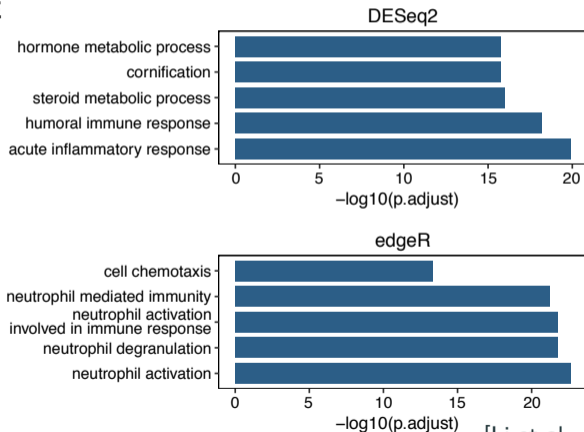
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## 2. Specification of a parametric model that does not fit data well

### Identifying differentially expressed genes (DEGs) from RNA-seq data

- False discoveries may mislead scientific conclusions

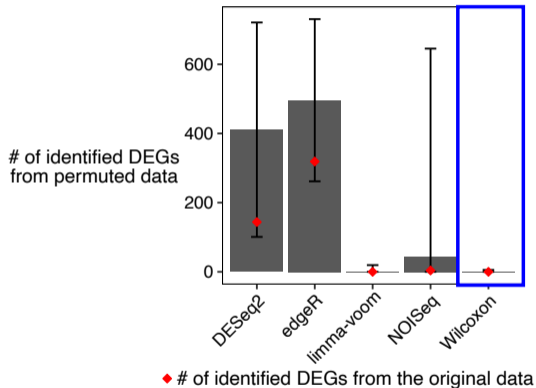


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## 2. Specification of a parametric model that does not fit data well

Method choice: popular bioinformatics tools vs. general statistical methods?



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Our recommendations for **large**-sample-sized data:

- **sanity check**: permutation
- consider **non-parametric** tests (e.g., Wilcoxon rank-sum test)

Short Report | [Open Access](#) | [Published: 15 March 2022](#)

### Exaggerated false positives by popular differential expression methods when analyzing human population samples

[Yumei Li](#), [Xinzhou Ge](#), [Fanglue Peng](#), [Wei Li](#) ✉ & [Jingyi Jessica Li](#) ✉

[Genome Biology](#) **23**, Article number: 79 (2022) | [Cite this article](#)

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— collaboration with Dr. Yumei Li in Dr. Wei Li's lab (UC Irvine)



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- **What if sample sizes are **small**?**

**Clipper** is a non-parametric option (to be elaborated)

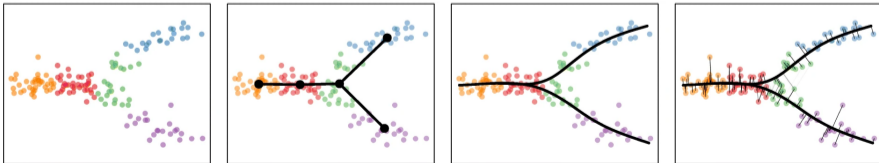
[Simons Big Data workshop in honor of Peter Bickel](#)



### 3. Treatment of inferred covariates as observed

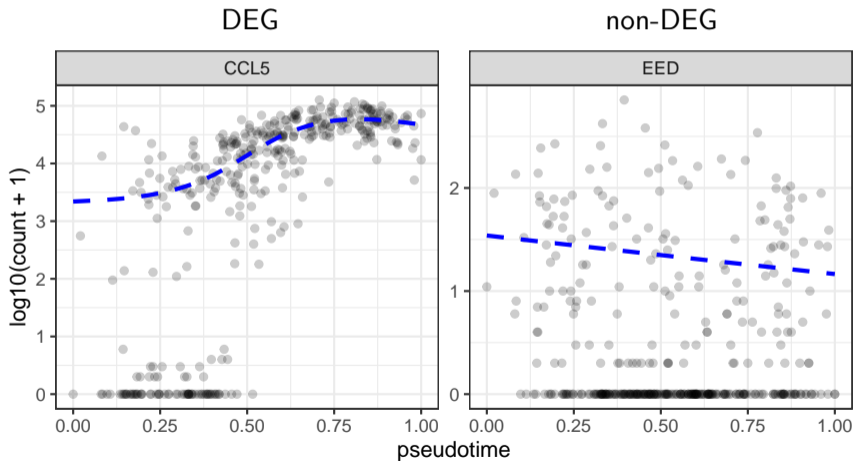
#### Example: identifying DEGs along pseudotime from single-cell RNA-seq data

- **Cell pseudotime**: a latent “temporal” variable that reflects a cell’s relative status among all cells
- **Pseudotime inference**: estimate the pseudotime of cells, i.e., order cells along a trajectory based on cells’ high-dimensional gene expression vectors
- Popular software:
  - Monocle3 [Trapnell *et al.*, *Nat Biotechnol*, 2014]; cited > 2.8K times
  - Slingshot [Street *et al.*, *BMC Bioinform*, 2018]; cited 700 times



### 3. Treatment of inferred covariates as observed

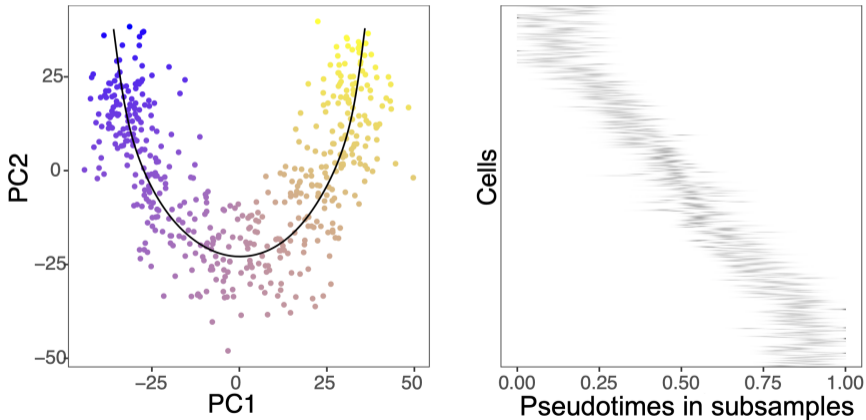
#### Identifying DEGs along inferred pseudotime from single-cell RNA-seq data



### 3. Treatment of inferred covariates as observed

#### Identifying DEGs along inferred pseudotime from single-cell RNA-seq data

- Cell pseudotime is inferred from the same data and thus **random**





### 3. Treatment of inferred covariates as observed

#### Identifying DEGs along inferred pseudotime from single-cell RNA-seq data

- However, existing methods treat cell pseudotime as an **observed covariate**



### 3. Treatment of inferred covariates as observed

#### Identifying DEGs along inferred pseudotime from single-cell RNA-seq data

- However, existing methods treat cell pseudotime as an **observed covariate**
- Our solution: **PseudotimeDE** considers the uncertainty of pseudotime inference

Method | [Open Access](#) | [Published: 29 April 2021](#)

#### **PseudotimeDE: inference of differential gene expression along cell pseudotime with well-calibrated $p$ -values from single-cell RNA sequencing data**

[Dongyuan Song](#) & [Jingyi Jessica Li](#) 

[Genome Biology](#) **22**, Article number: 124 (2021) | [Cite this article](#)

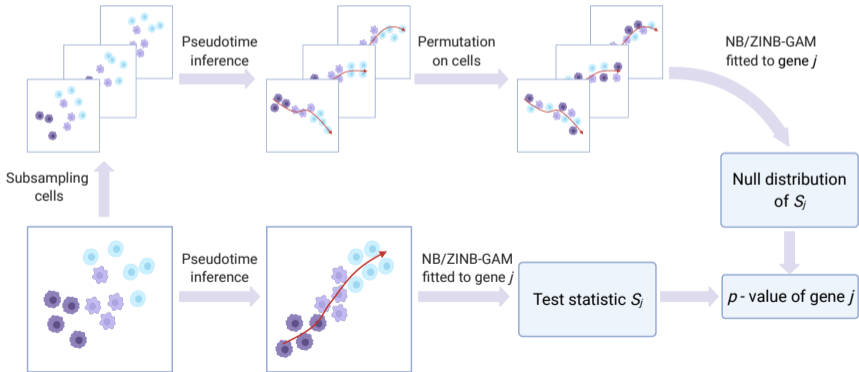
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#### Identifying DEGs along inferred pseudotime from single-cell RNA-seq data

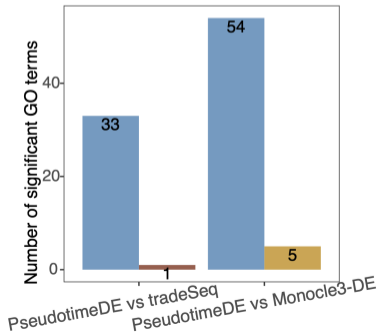
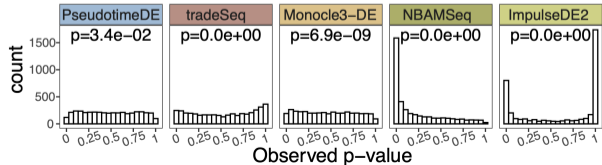
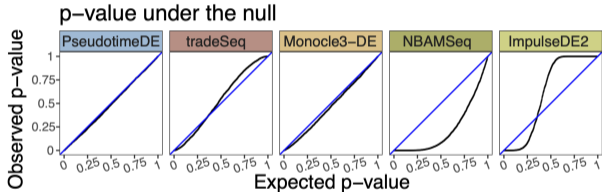
- **PseudotimeDE** generates well-calibrated p-values for FDR control & uses a **generalized additive model (GAM)** to achieve good power



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#### Identifying DEGs along inferred pseudotime from single-cell RNA-seq data

##### PseudotimeDE limitations

- **computational time**: high-resolution p-values require  $> 10^3$  rounds of (subsampling + pseudotime inference + permutation)



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Q: how to reduce the number of rounds while still achieving FDR control?

A: **Clipper**



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A: **Clipper**

- **complete null**: what if cells do not follow a trajectory

Q: how to generate the null cells?

A: **simulator scDesign3**





### 3. Treatment of inferred covariates as observed

Identifying DEGs along inferred **pseudotime** from single-cell RNA-seq data

- **PseudotimeDE**

Identifying DEGs between inferred **cell clusters** from single-cell RNA-seq data

- **ClusterDE** (cell clustering + DEG identification between cell clusters)
  - existing methods assume Gaussian distributions
    - TN test [Zhang, Kamath, and Tse, *Cell Syst*, 2019]
    - clusterpval [Gao, Bien, and Witten, *arXiv*, 2020]



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**Our proposal: Clipper + scDesign3**

— inspired by

gap statistic [Hastie, Tibshirani, and Walther, *JRSSB*, 2002]

knockoffs [Barber and Candès, *Ann Stat*, 2015]



### Three common causes of ill-posed p-values

1. Formulation of a **two-sample test** as a one-sample test
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### **Clipper**: p-value-free FDR control for genomics feature screening

— using FDR control procedure from [Barber and Candès, *Ann Stat*, 2015]



# Clipper: p-value-free FDR control for genomics feature screening



- **NO requirement of**
  - high-resolution p-values
  - parametric distributions
  - large sample sizes
- **Foundation: knockoffs**
- **Two components**
  - **contrast scores**
  - **cutoff**

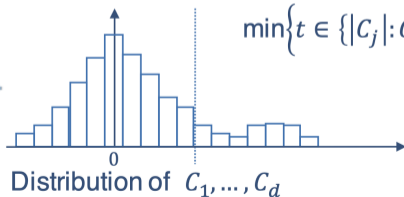
**Goal:** marginal screening for **interesting** features

$d$  features

FDR threshold  $q$

Contrast scores

$C_1$   
 $\vdots$   
 $C_d$



Contrast score cutoff

$$\min \left\{ t \in \{|C_j| : C_j \neq 0\} : \frac{1 + \#\{j : C_j \leq -t\}}{\#\{j : C_j \geq t\} \vee 1} \leq q \right\}$$



# Clipper: p-value-free FDR control for genomics feature screening

Key: **contrast score** construction

example	target data	null data
ChIP-seq peak calling (1 vs. 1)	experimental condition	background condition
RNA-seq DEG identification	actual data	permuted data
<b>PseudotimeDE &amp; ClusterDE</b>	actual data	<b>scDesign3</b> simulated data

**Contrast score** of feature  $j = 1, \dots, d$ , the

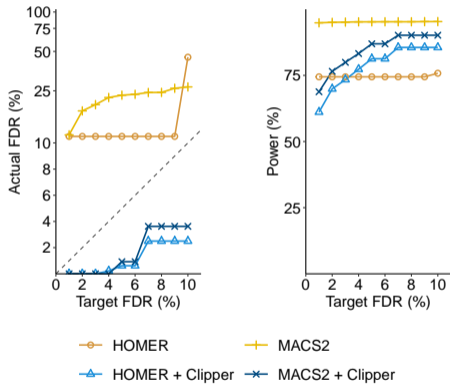
$$C_j := t(\text{target data}) - t(\text{null data}),$$

where  $t(\cdot)$  is a summary statistic — can be a **complex pipeline**

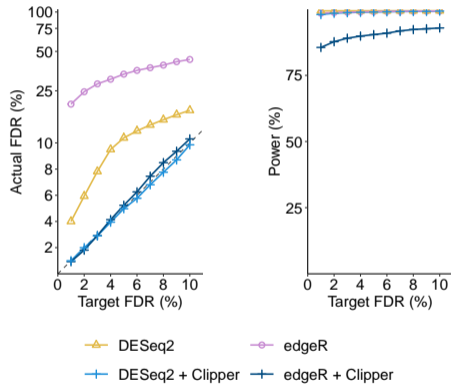


# Clipper rectifies FDR control

## ChIP-seq peaking calling

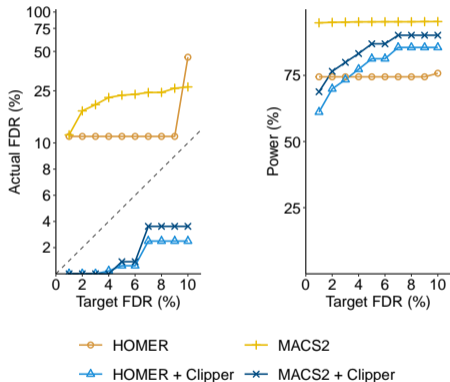


## RNA-seq DEG identification

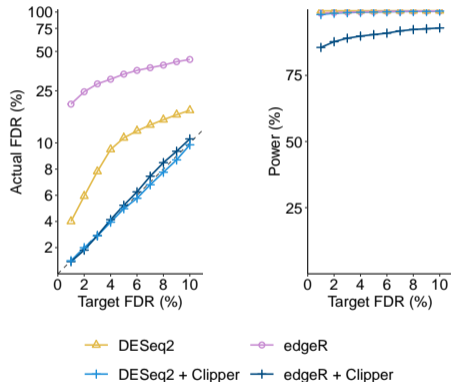


# Clipper rectifies FDR control

## ChIP-seq peaking calling



## RNA-seq DEG identification



Q: how to generate null data to construct contrast scores for PseudotimeDE and ClusterDE?



## Three common causes of ill-posed p-values

1. Formulation of a **two-sample test** as a one-sample test
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**Clipper**: a p-value-free FDR control framework

**scDesign3**: an omnibus single-cell omics simulator



# scDesign2: a probabilistic single-cell gene expression data simulator

A multi-gene probabilistic model **per cell type**

- Each gene  $\sim$  count distribution  $\in$  {Poisson, negative binomial, ZIP, ZINB}
- Gene correlations estimated via **Gaussian copula**

Method | [Open Access](#) | [Published: 25 May 2021](#)

## scDesign2: a transparent simulator that generates high-fidelity single-cell gene expression count data with gene correlations captured

[Tianyi Sun](#), [Dongyuan Song](#), [Wei Vivian Li](#)  & [Jingyi Jessica Li](#) 

[Genome Biology](#) **22**, Article number: 163 (2021) | [Cite this article](#)

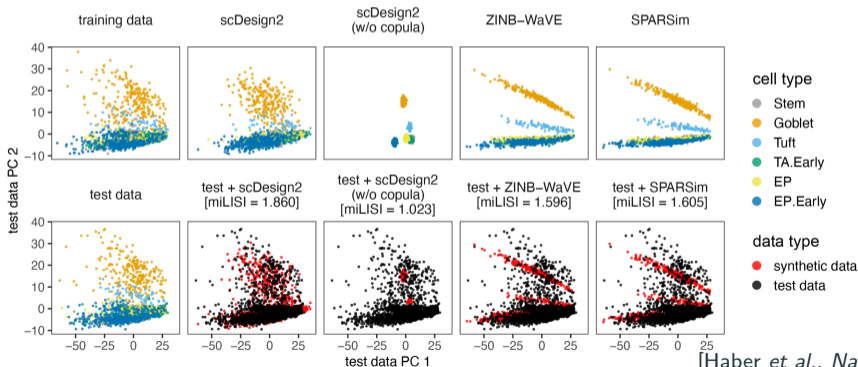
**5144** Accesses | **8** Citations | **31** Altmetric | [Metrics](#)



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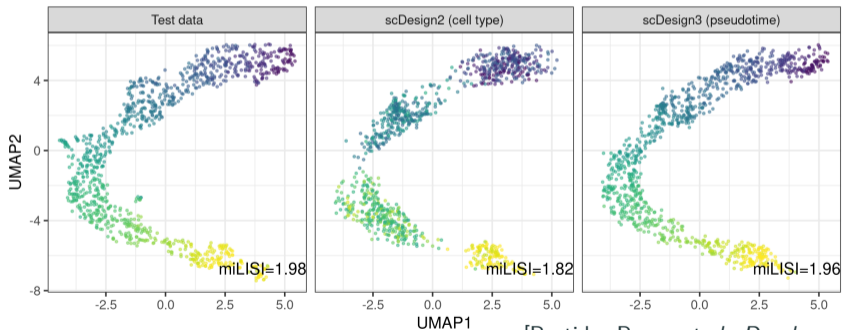


[Haber *et al.*, *Nature*, 2017]

# scDesign3: an omnibus single-cell & spatial omics simulator

- **Cell states:** continuous trajectory & discrete cell types
- **Feature modalities:** RNA, ATAC, protein, spatial coordinates, etc.
- **Model selection by likelihood:** vine copula [Joe and Kurowicka's book, 2011]

Example: continuous trajectory (pancreatic cell differentiation)



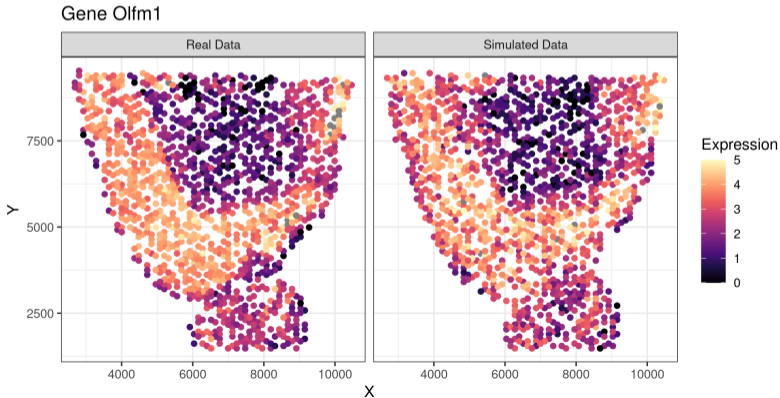
[Bastidas-Ponce *et al.*, *Development*, 2019]



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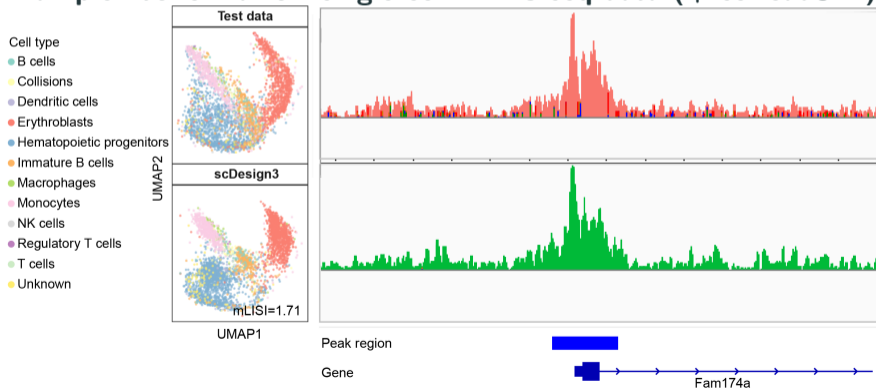
**Example: spatial data (brain region measured by 10X Visium)**



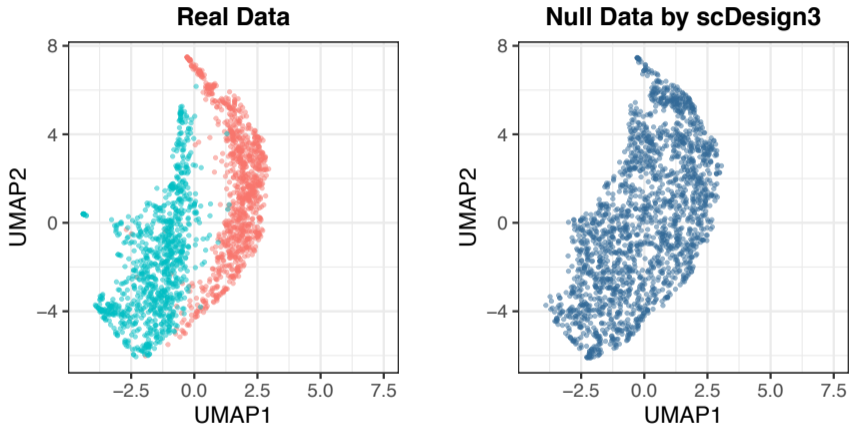
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## Example: bone marrow single-cell ATAC-seq data (+ scReadSim)



# ClusterDE: scDesign3 for null data generation (preliminary)



Cell\_Type ● naive.cytotoxic ● regulatory.t

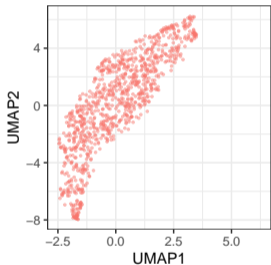
[Zheng *et al.*, *Nat Commun*, 2017]



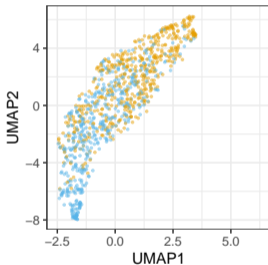
# ClusterDE: Clipper + scDesign3 (preliminary)

Complete null case: no cell clusters

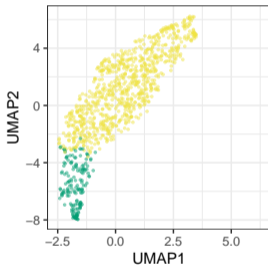
Real Data



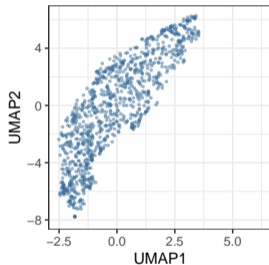
Seurat Clustering



Kmeans Clustering



Null Data by scDesign3



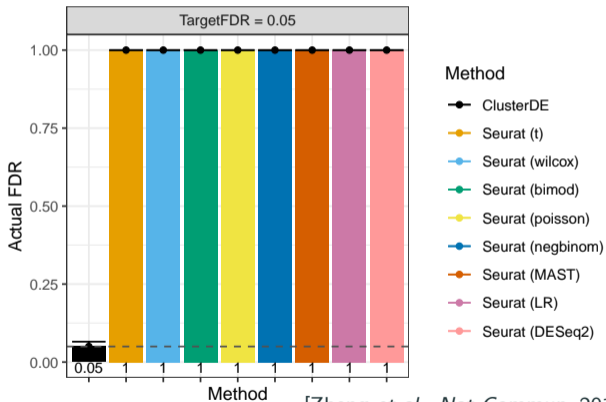
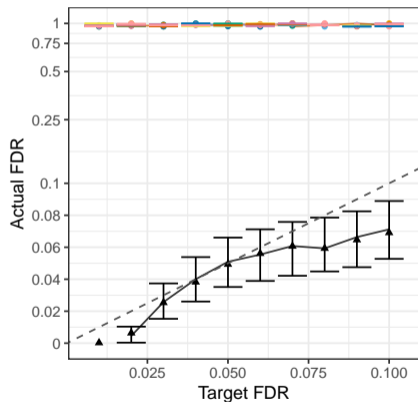
[Zheng et al., Nat Commun, 2017]





# ClusterDE: Clipper + scDesign3 (preliminary)

Complete null case: no cell clusters



[Zheng et al., Nat Commun, 2017]



# Summary: p-values are easily ill-posed in genomics data analysis

Q: should a scientific question be formulated as multiple testing?

## Patterns



Perspective

# Statistical Hypothesis Testing versus Machine Learning Binary Classification: Distinctions and Guidelines

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## Summary: p-values are easily ill-posed in genomics data analysis

Q: should a scientific question be formulated as multiple testing?

If YES, three common causes of ill-posed p-values

1. Formulation of a **two-sample test** as a one-sample test
  - ChIP-seq peak calling
2. Specification of a **parametric model** that does not fit data well
  - RNA-seq DEG identification
3. Treatment of **inferred** covariates as observed
  - single-cell RNA-seq PseudotimeDE & ClusterDE



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**Clipper**: a p-value-free FDR control framework

**scDesign3**: an omnibus single-cell & spatial omics simulator

– fair benchmarking of computational tools (> 1000 at [www.scrna-tools.org](http://www.scrna-tools.org))



# Summary: relevant publications

Short Report | [Open Access](#) | [Published: 15 March 2022](#)

## Exaggerated false positives by popular differential expression methods when analyzing human population samples

[Yumei Li](#), [Xinzhou Ge](#), [Fanglue Peng](#), [Wei Li](#)  & [Jingyi Jessica Li](#) 

[Genome Biology](#) **23**, Article number: 79 (2022) | [Cite this article](#)

14k Accesses | 185 Altmetric | [Metrics](#)

Method | [Open Access](#) | [Published: 29 April 2021](#)

## PseudotimeDE: inference of differential gene expression along cell pseudotime with well-calibrated $p$ -values from single-cell RNA sequencing data

[Dongyuan Song](#) & [Jingyi Jessica Li](#) 

[Genome Biology](#) **22**, Article number: 124 (2021) | [Cite this article](#)

7705 Accesses | 4 Citations | 29 Altmetric | [Metrics](#)

Method | [Open Access](#) | [Published: 11 October 2021](#)

## Clipper: $p$ -value-free FDR control on high-throughput data from two conditions

[Xinzhou Ge](#), [Yiling Elaine Chen](#), [Dongyuan Song](#), [MeiLu McDermott](#), [Kyla Woyschner](#), [Antigoni Manousopoulou](#), [Ning Wang](#), [Wei Li](#), [Leo D. Wang](#) & [Jingyi Jessica Li](#) 

[Genome Biology](#) **22**, Article number: 288 (2021) | [Cite this article](#)

6169 Accesses | 4 Citations | 52 Altmetric | [Metrics](#)

Method | [Open Access](#) | [Published: 25 May 2021](#)

## scDesign2: a transparent simulator that generates high-fidelity single-cell gene expression count data with gene correlations captured

[Tianyi Sun](#), [Dongyuan Song](#), [Wei Vivian Li](#)  & [Jingyi Jessica Li](#) 

[Genome Biology](#) **22**, Article number: 163 (2021) | [Cite this article](#)

5144 Accesses | 8 Citations | 31 Altmetric | [Metrics](#)



### PCA outperforms popular hidden variable inference methods for QTL mapping

 Heather J. Zhou,  Lei Li,  Yumei Li,  Wei Li,  Jingyi Jessica Li

doi: <https://doi.org/10.1101/2022.03.09.483661>

“These results may come as a surprise to some, given the nearly uncontested status that *method A* has achieved within the community, but sadly they reflect the fact that computational biology methods can rise to fame almost **by accident rather than by sound statistical arguments.**”



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**Dr. Yumei Li**  
(Collaborator  
postdoc  
@ UCI)  
DE genes



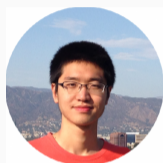
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(Postdoc)  
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**Dr. Yiling Elaine Chen**  
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**Tianyi Sun**  
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scDesign2



**Dongyuan Song**  
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PseudotimeDE  
scDesign3

