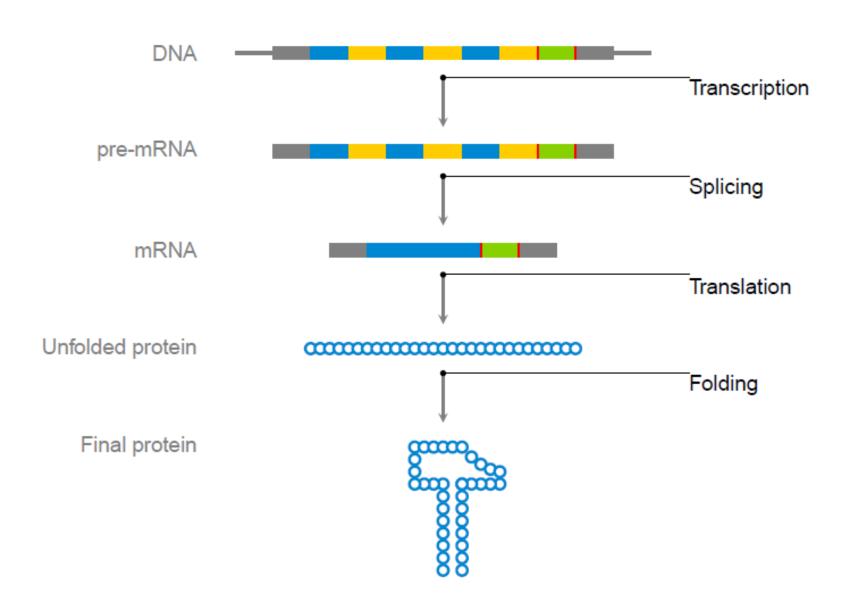
# Molecular errors, cryptic sequences, and evolvability

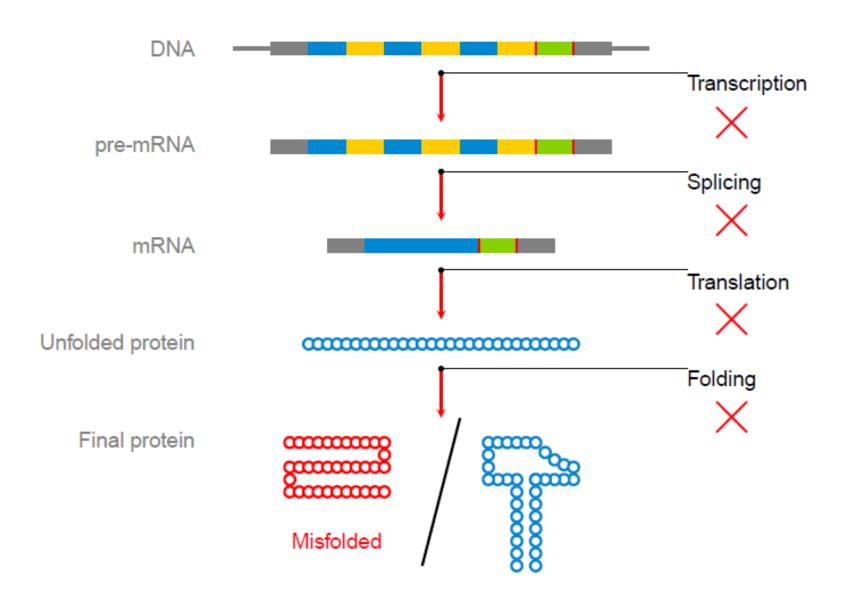
Joanna Masel

Ecology & Evolutionary Biology, University of Arizona

### Gene expression



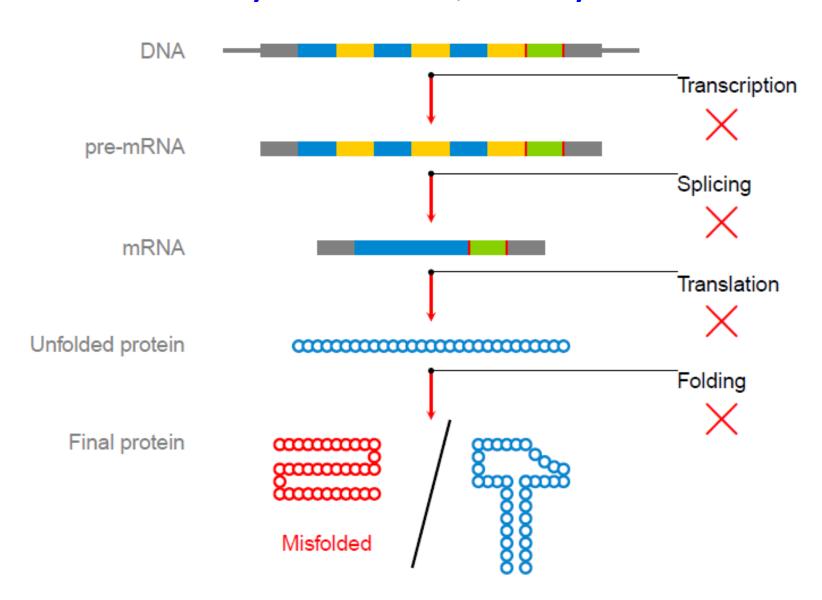
### Errors can occur at any stage



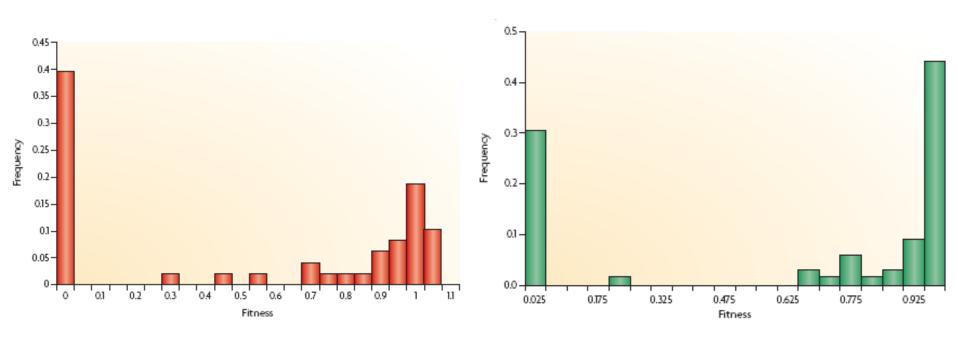
### Outline

- 1. Evolution of error rates under a speed vs. accuracy tradeoff
- 2. Molecular errors pre-screen future variants, and so promote evolvability
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# Consequences of errors are either bad or relatively harmless, rarely in between



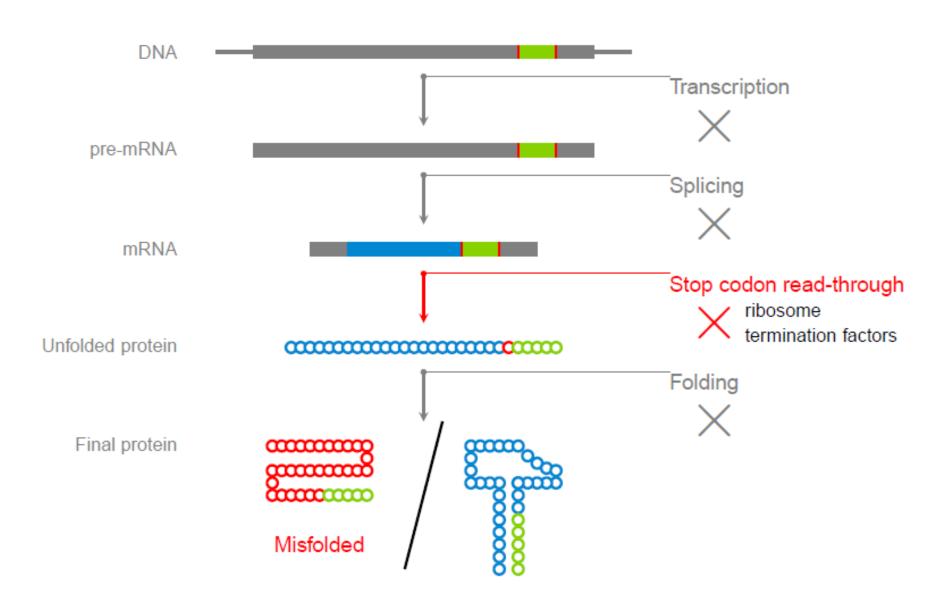
# Distribution of fitness effects of new mutations



vesicular stomatic virus

yeast

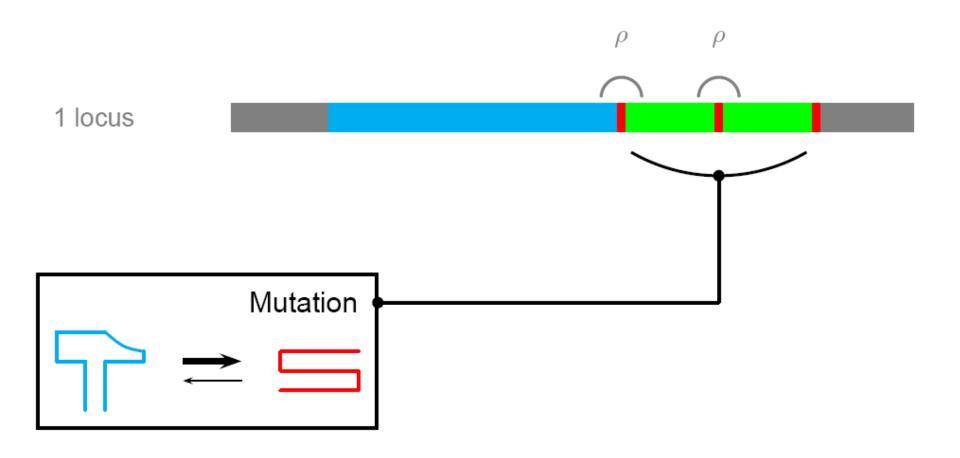
# Stop codon readthrough: case study of molecular errors



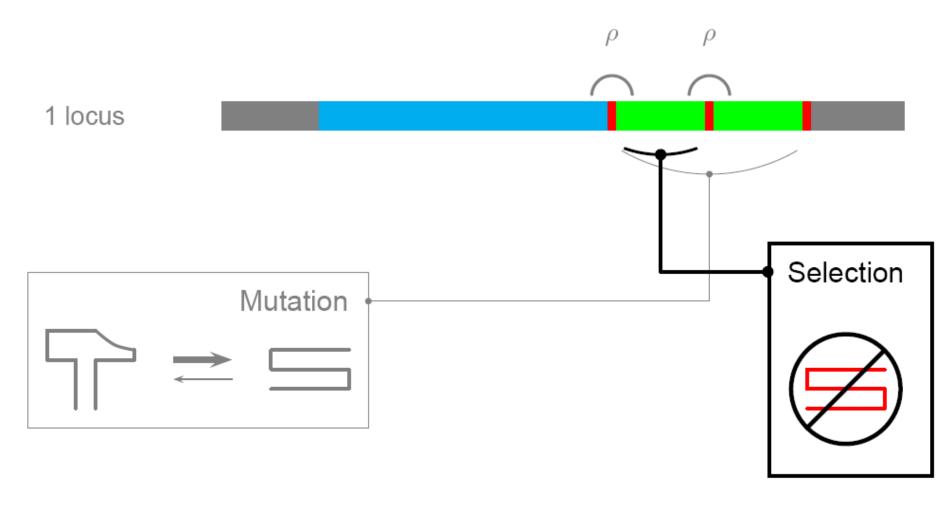
### Readthrough at error rate $\rho$



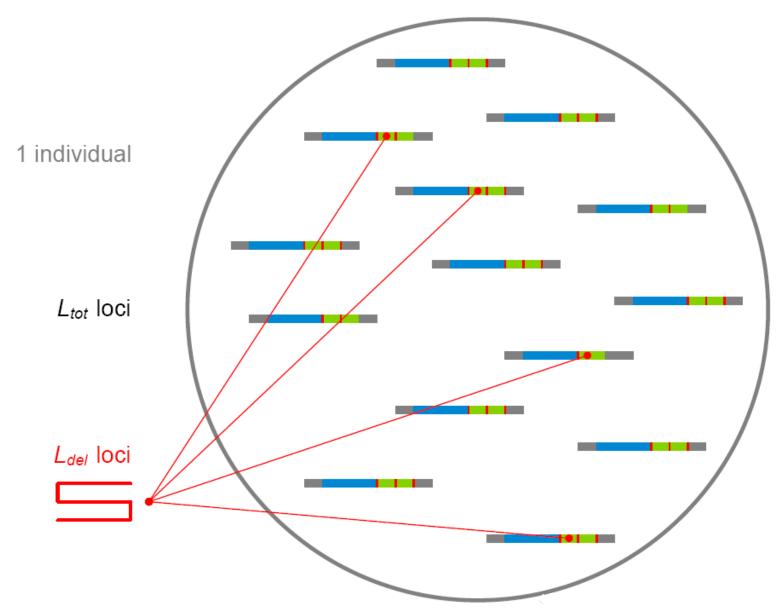
### Mutation bias favors misfolding



# Selection for a stable fold even after a readthrough error

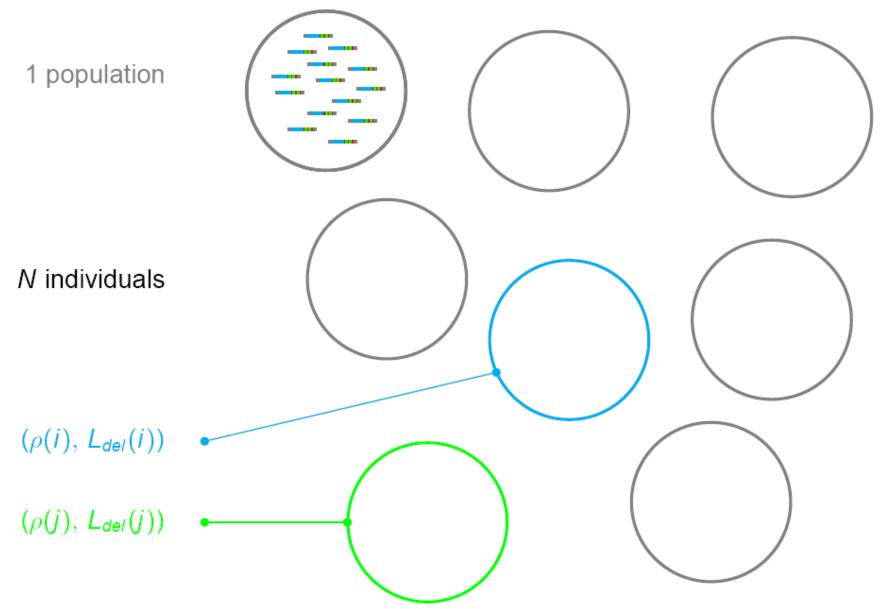


# Readthrough errors happen at many loci. Some are sensitive.

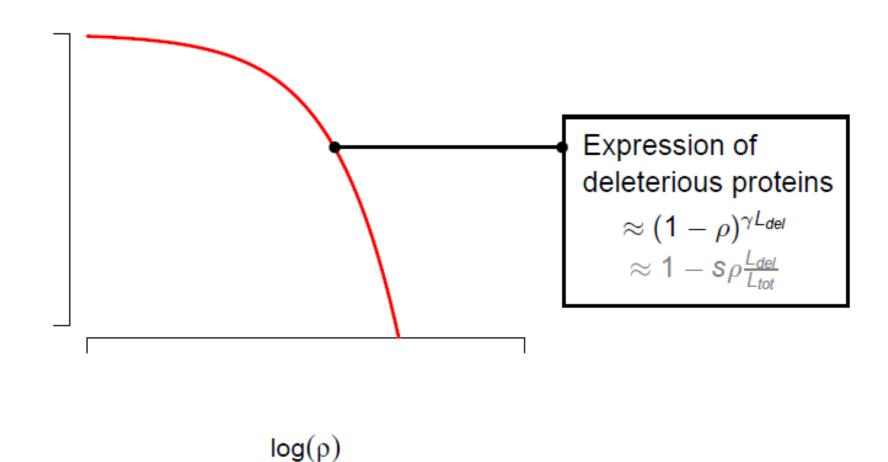


### Individual genotype

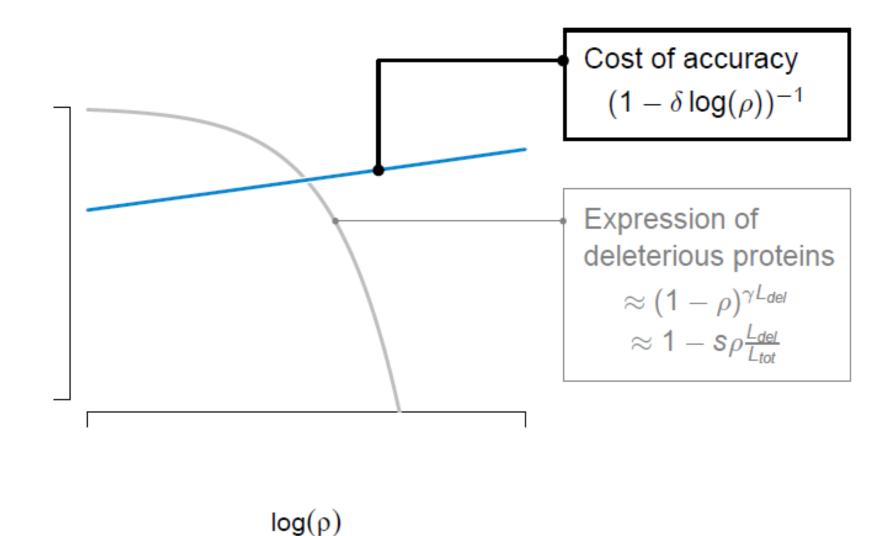
= error rate, #sensitive loci



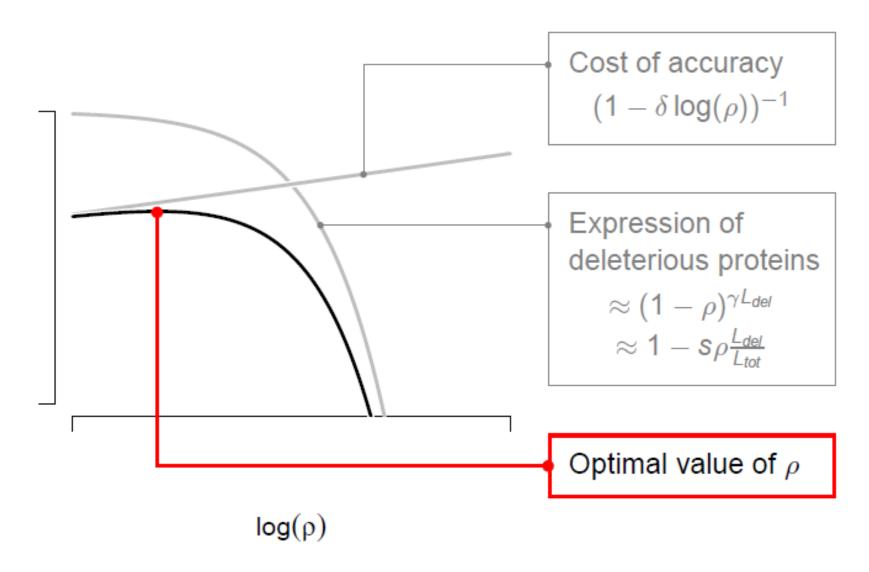
### Costs and benefits of proofreading



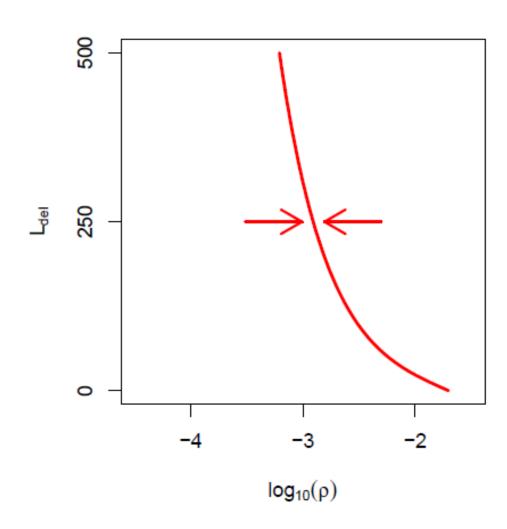
### Costs and benefits of proofreading



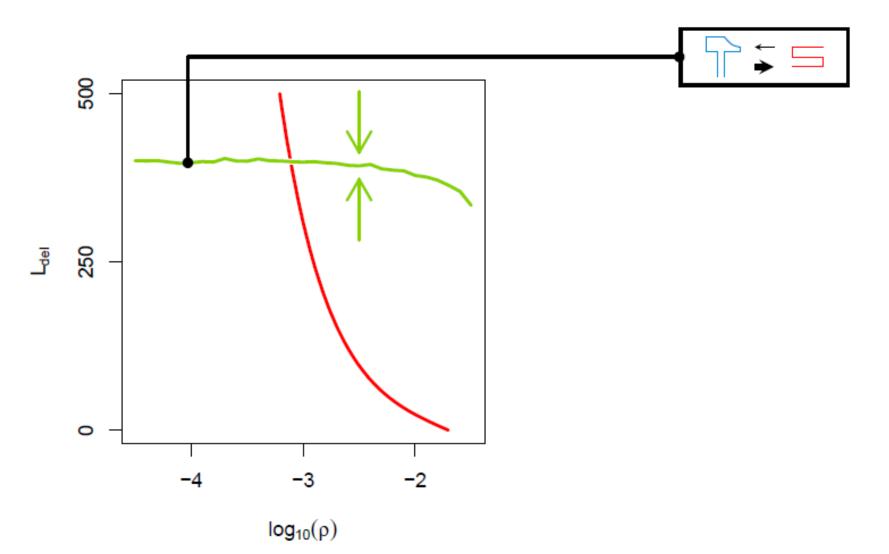
### Costs and benefits of proofreading



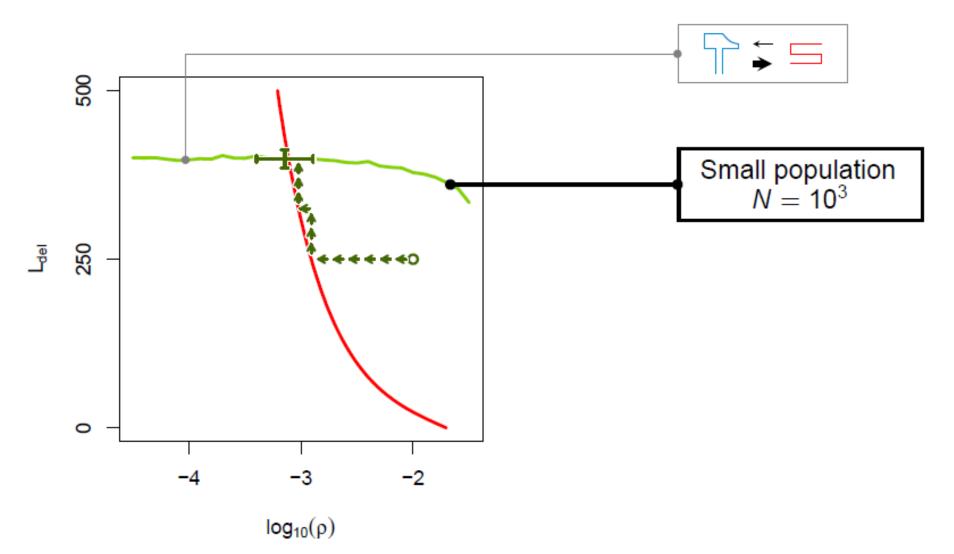
## Coevolution of $\rho$ and $\textbf{L}_{\text{del}}$



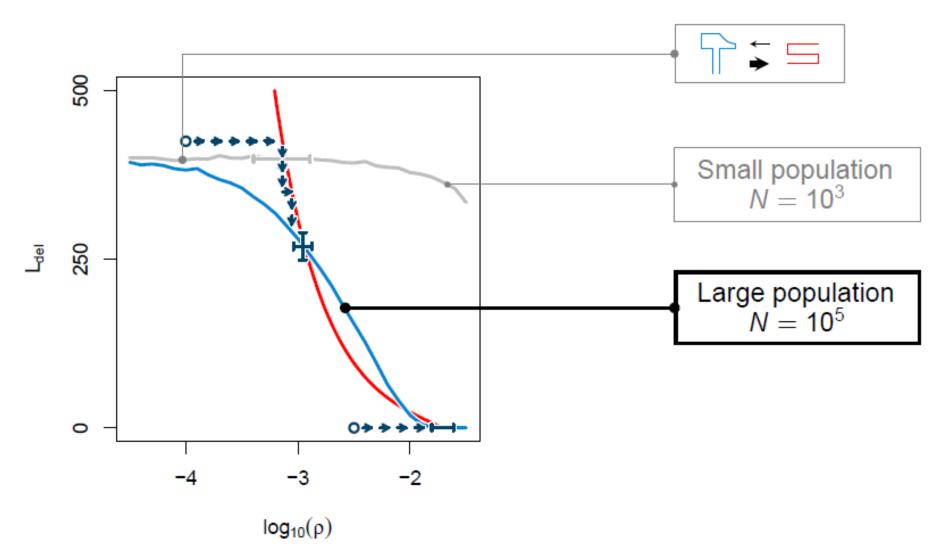
## Coevolution of $\rho$ and $\textbf{L}_{\text{del}}$



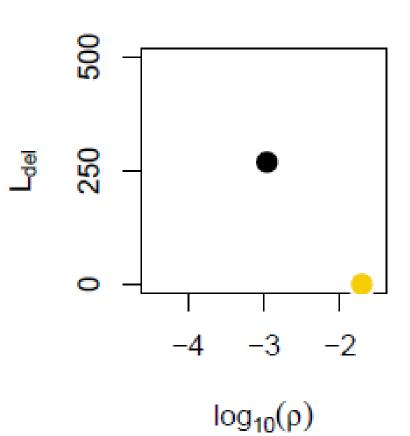
### Coevolution of $\rho$ and $\textbf{L}_{\text{del}}$



#### Two attractors in large populations



#### Two strategies are quite different

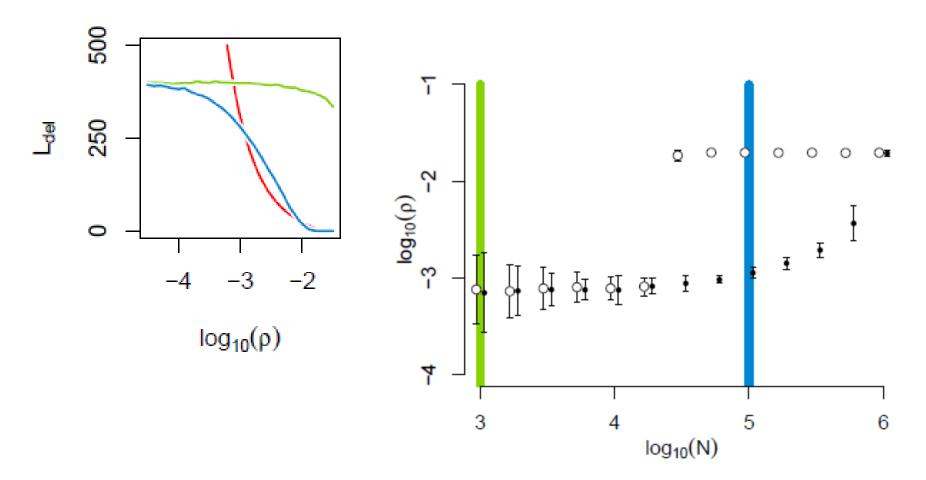


#### 2 strategies:

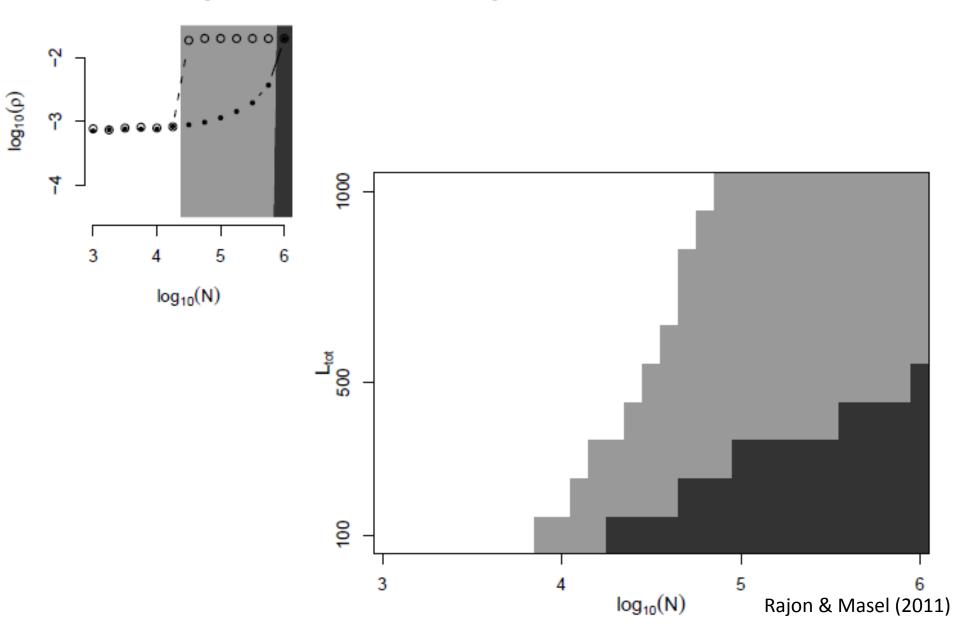
- allowing deleterious sequences, but hiding them
- eliminating deleterious sequence by expressing them

or ●?

# Two attractors for a range of population sizes (i.e. range of limits to weak selection)



### Larger bistable range with more loci



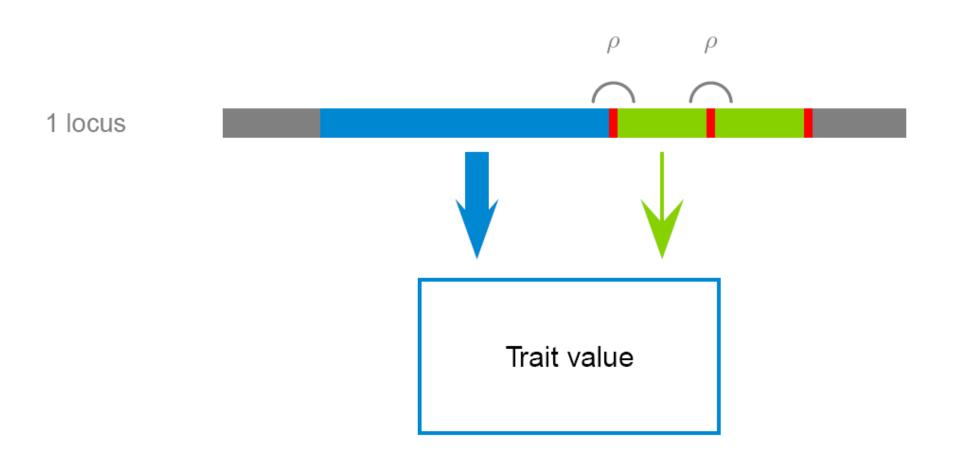
# Model applies to many kinds of molecular errors

Error	Global solution	Local solution
Stop codon readthrough	Accurate ribosome & release factors	Benign 3'UTR

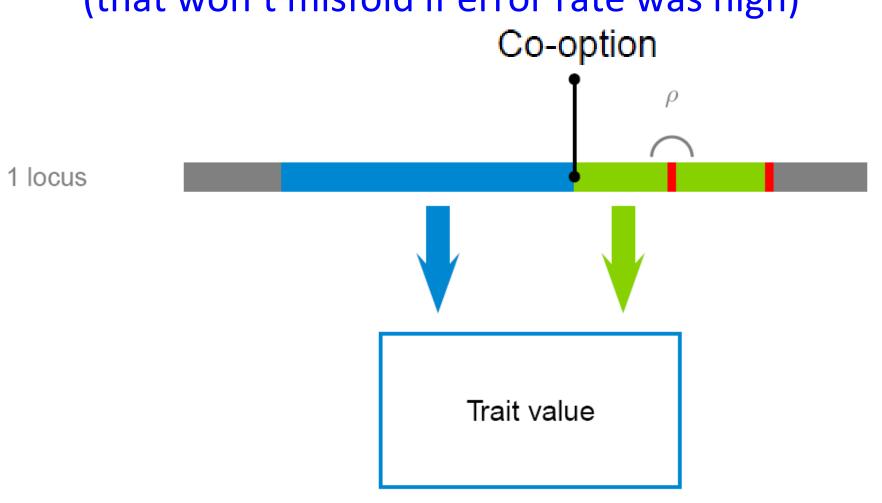
### Outline

- 1. Evolution of error rates under a speed vs. accuracy tradeoff
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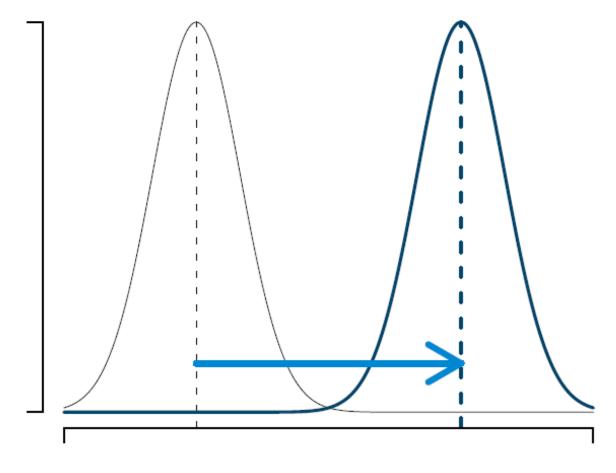
# Effect on quantitative trait proportional to expression



# Point mutation in stop codon → full expression of previously cryptic sequence (that won't misfold if error rate was high)



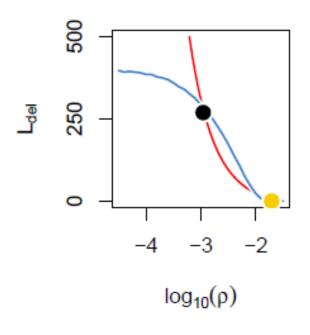
# Environmental change in optimal trait value

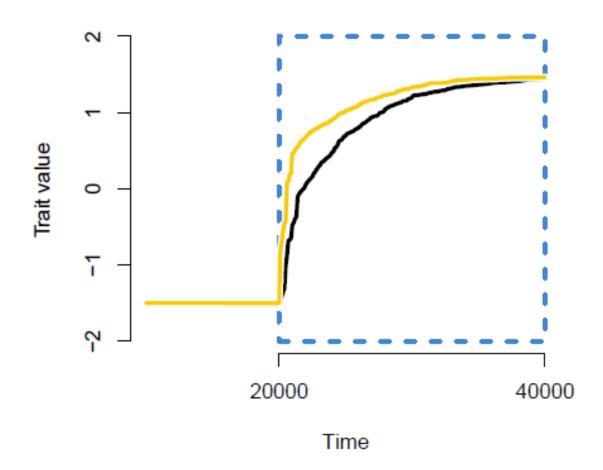


Fitness

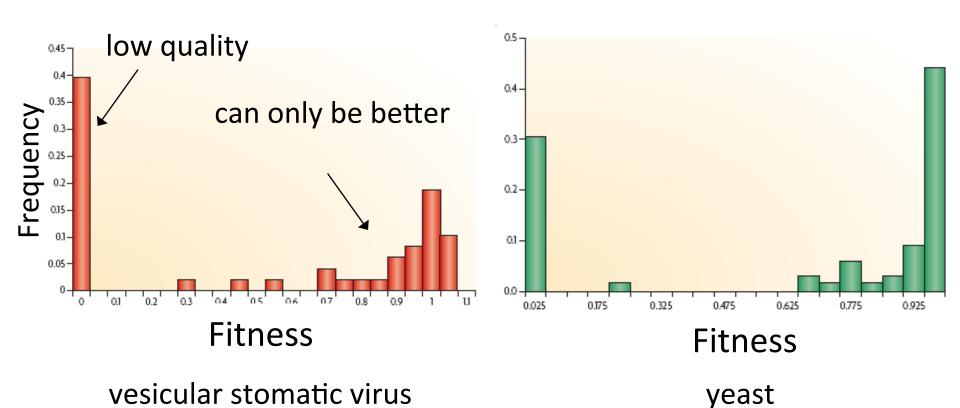
Trait value

# Populations with high error rates evolve faster



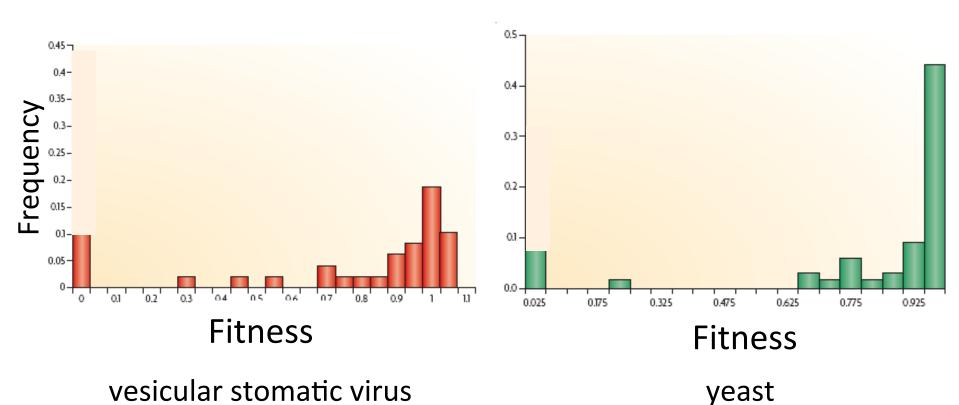


#### **New mutations**



Eyre-Walker & Keightley 2007

### **Cryptic variants**



vesicular stomatic virus

**Pre-adapting selection** 

Masel 2006, Rajon & Masel 2011

# Evolvability comes from tapping into cryptic variants

- Molecular errors in the present mimic mutations in the future
- Strongly deleterious sequences are pre-purged in favor of benign ones
- Benign sequences are co-optable for adaptation

# Benefits go to any "high error" locally benign cryptic sequences

#### More examples

- Promiscuous enzyme activities
- Rare protein-protein interactions (PPIs) that lose crypticity when proteins see each other more often

Aside: "cryptic" PPIs (deliberately bad Y2H data) are biologically meaningful

They predict gene noise and plasticity better than "real" PPIs (best practice affinity capture mass spec)

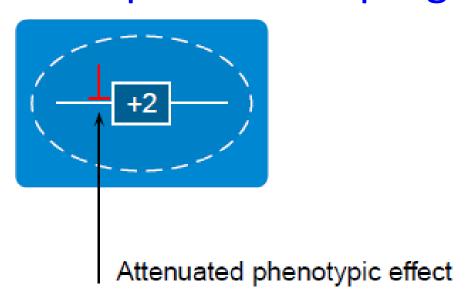
"Stickiness" trumps "hubness"

### Outline

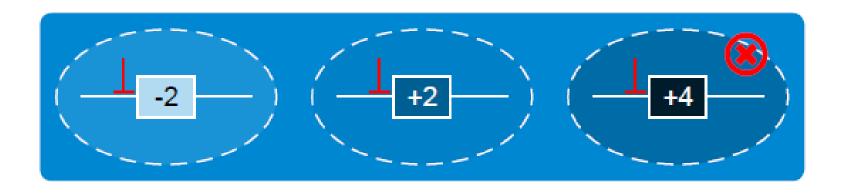
- Evolution of error rates under a speed vs. accuracy tradeoff
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# Let's look at cryptic sequences with and without genetic diversity

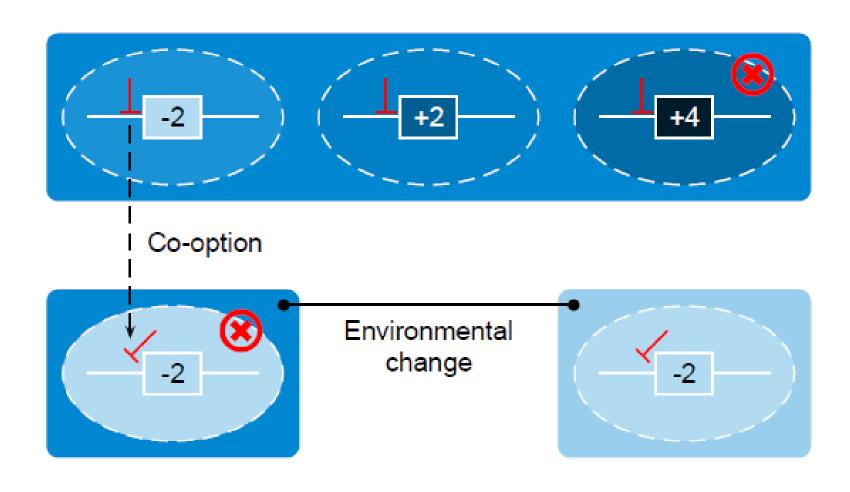
Consider only benign sequences, with different phenotypic effect sizes (i.e. in parameter regime where misfolded cryptic sequences are purged)



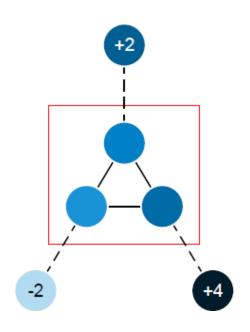
# Relaxed selection → cryptic genetic diversity



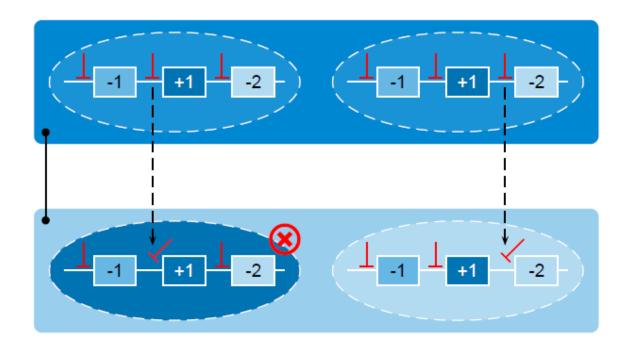
## Co-opted variants can be adaptive in a new environment



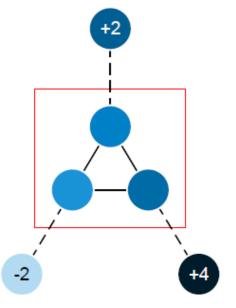
### Genotype space / neutral network



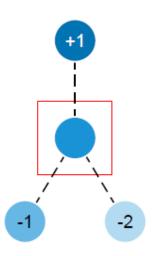
# Multiple cryptic loci provide more adaptive options, even in the absence of genetic diversity across population



## Two ways to access more novel phenotypes: genetic polymorphism or neighborhood richness

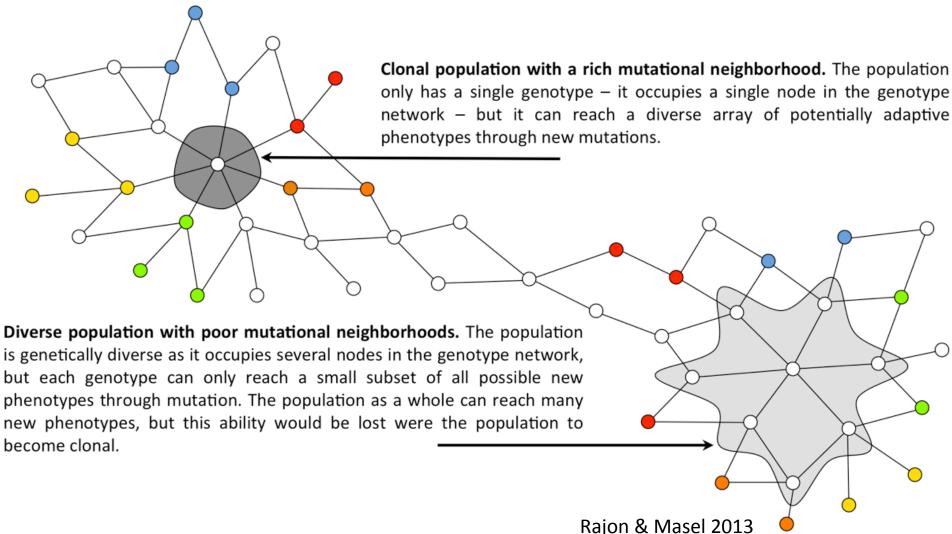


1 locus, 3 genotypes, each accessing one new phenotype



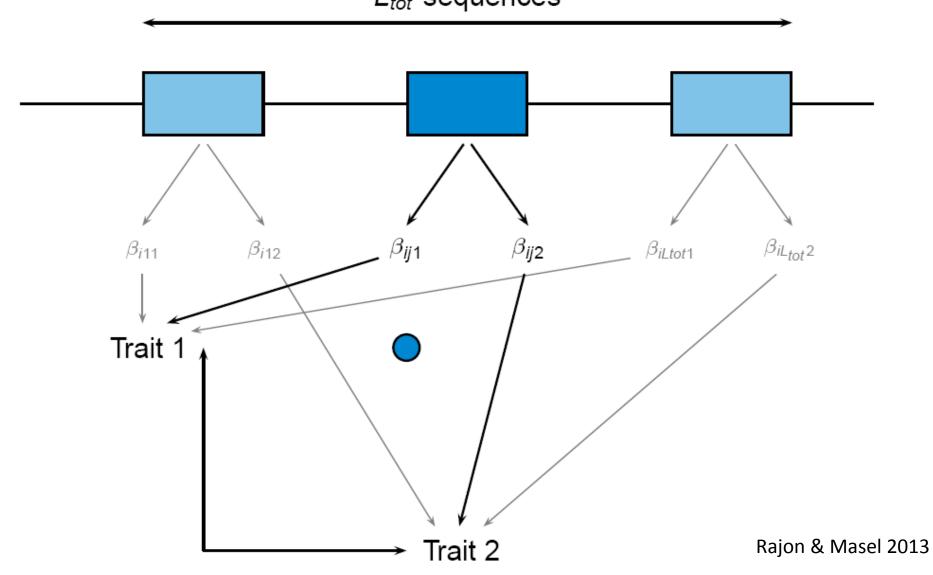
3 loci, 1 genotype can access 3 phenotypes

## Two ways to access more novel phenotypes: genetic polymorphism or neighborhood richness

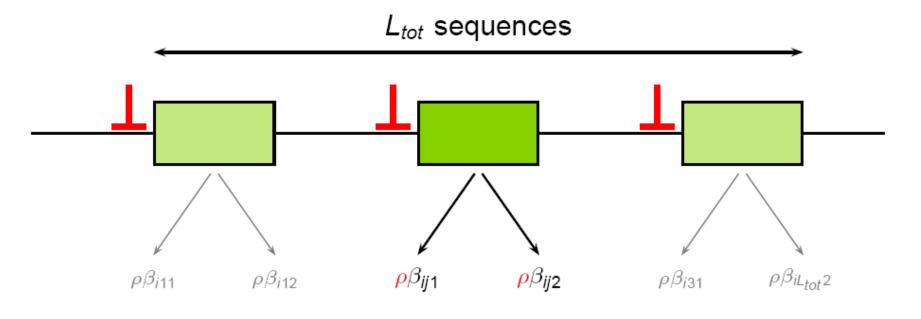


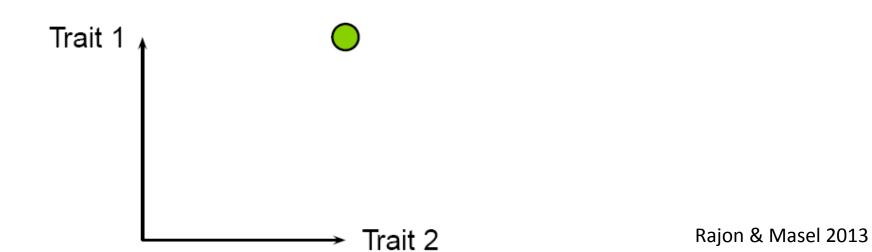
## Each cryptic sequence affects multiple traits

*L*<sub>tot</sub> sequences

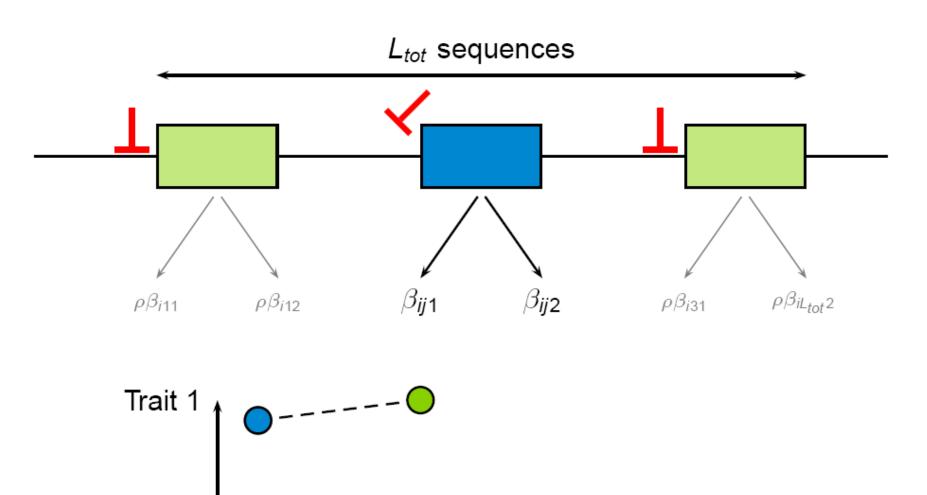


### Effects are dampened while cryptic



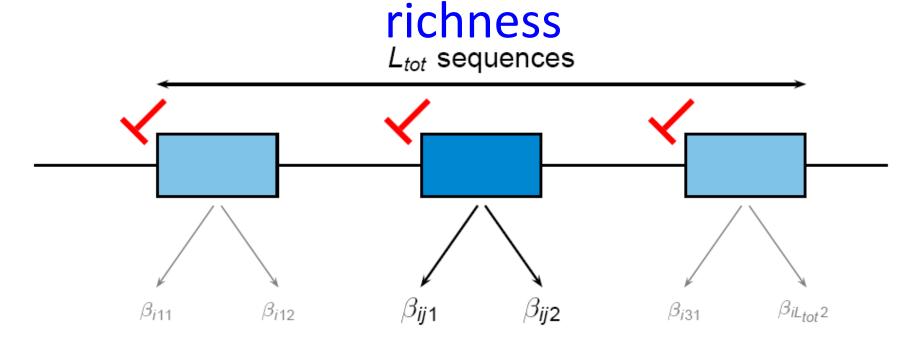


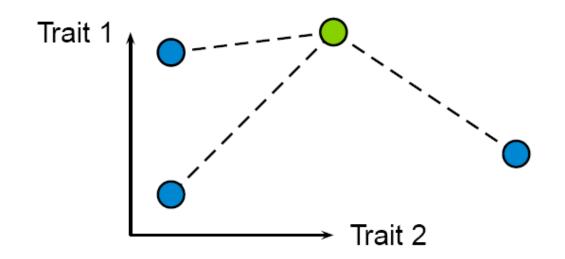
### During co-option, crypticity is lost



Trait 2

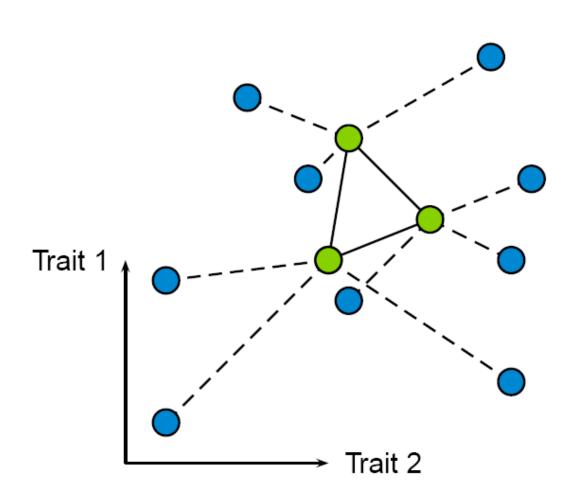
### Multiple sequences define neighborhood





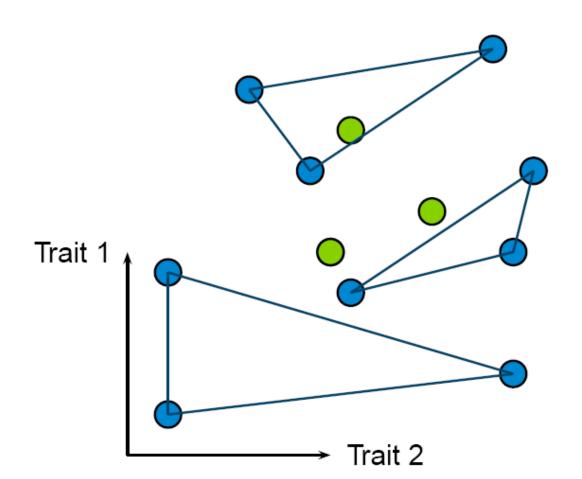
Rajon & Masel 2013

## Multiple genotypes increase accessible phenotypes still further



## Quantify phenotypic diversity due to neighborhood richness

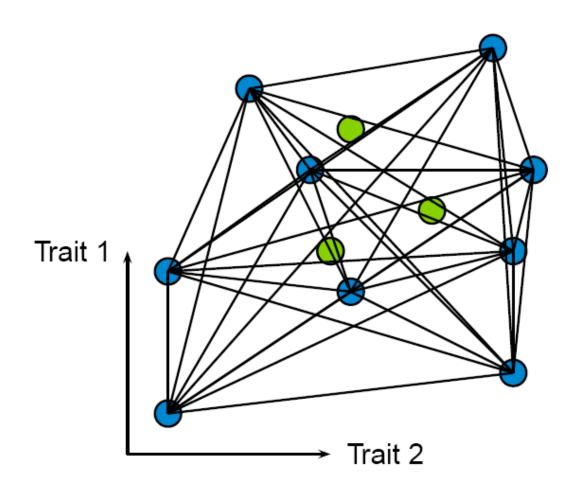
d<sub>G</sub>: mean distance between individuals with the same initial genotype



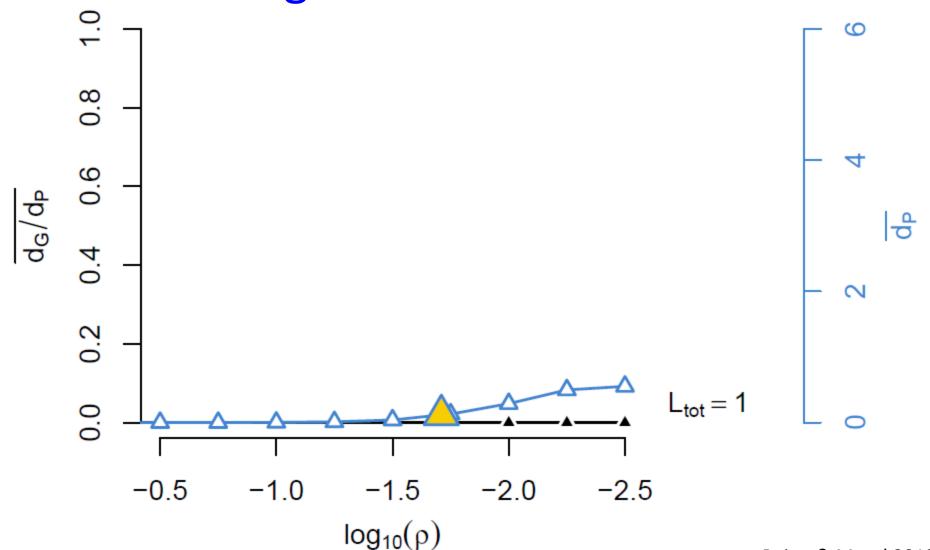
### Compare to total phenotypic diversity

d<sub>G</sub>: mean distance between individuals with the same initial genotype

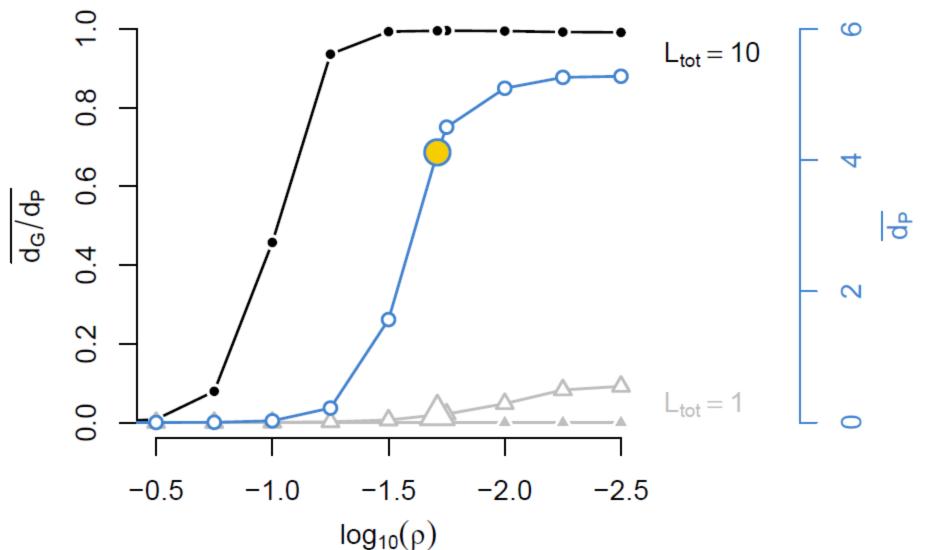
*d<sub>P</sub>*: mean distance between two individuals in the population



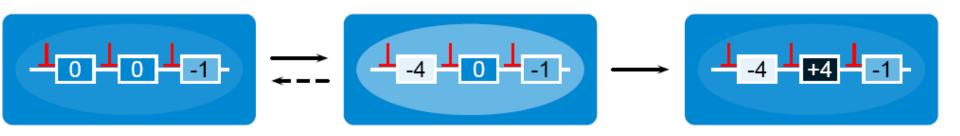
## With one locus, all genetic diversity, no neighborhood richness



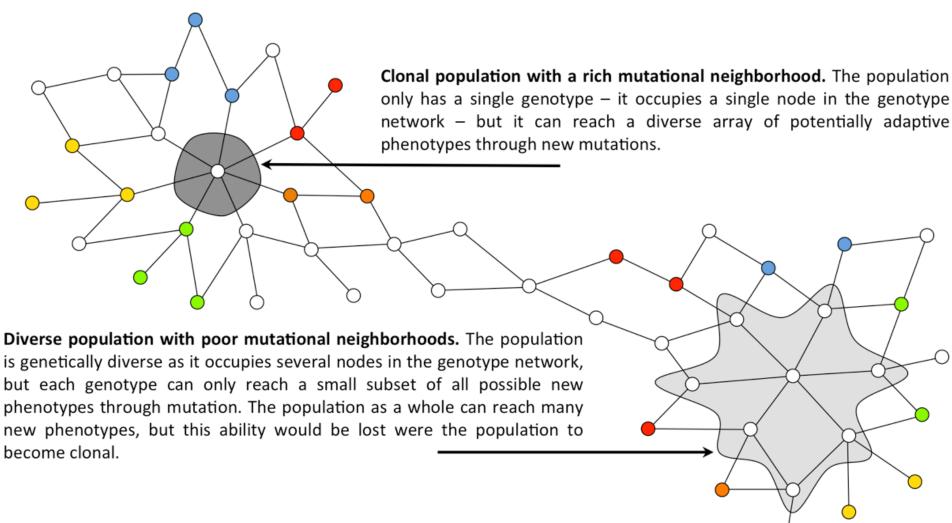
## With 10 loci, more phenotypic diversity, dominated by neighborhood richness



## Compensatory evolution drives high neighborhood richness



## "Spread" across a genotype space is not required for the high evolvability of polygenic traits in asexuals



Rajon & Masel 2013

### What do we need for evolvability?

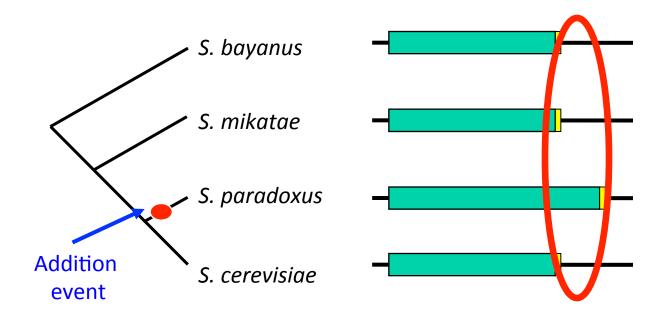
- A minimum level of selection on cryptic sequences, to purge the misfolded options
- Selection as weak as possible above that minimum, to allow maximum compensatory evolution

 This balance is exactly what we get in one attractor of our speed vs. accuracy model!

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## Stop codon readthrough can be coopted for de novo C-terminal pieces of genes



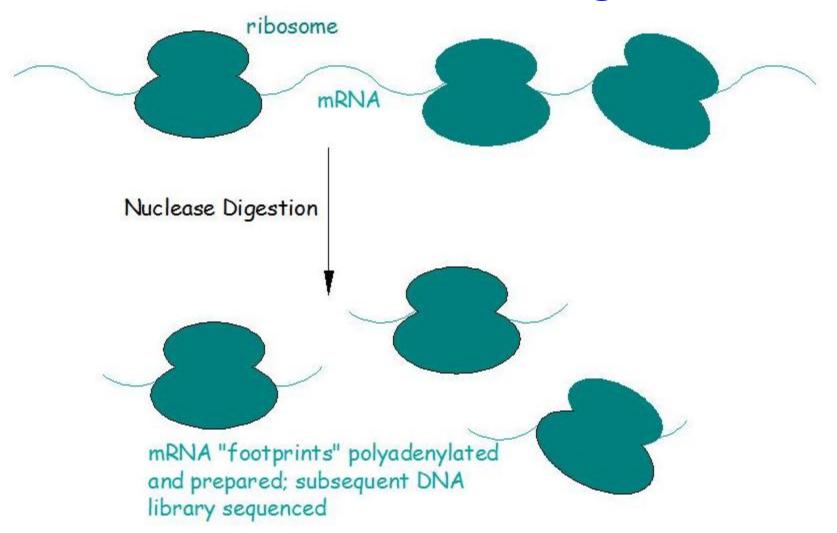
- Conversion of non-coding to coding confirmed by homologous phylogenetic comparisons
  - 75 events in Saccharomyces
  - 67 events in mouse/rat

### Complete genes evolve *de novo* too. How is this possible?

- 1. Accidental, low level transcription, transcript rapidly degraded
- 2. Transcript escapes degradation
- Transcript occasionally exported to cytoplasm, where it associates with ribosomes and "accidental" ORFs may be translated at low levels
- 4. New, functional coding gene

- Errors at each stage give a "preview" of the next one, allowing pre-adaptation to occur
- We tested whether penultimate stage 3 is common

### Ribosome Profiling

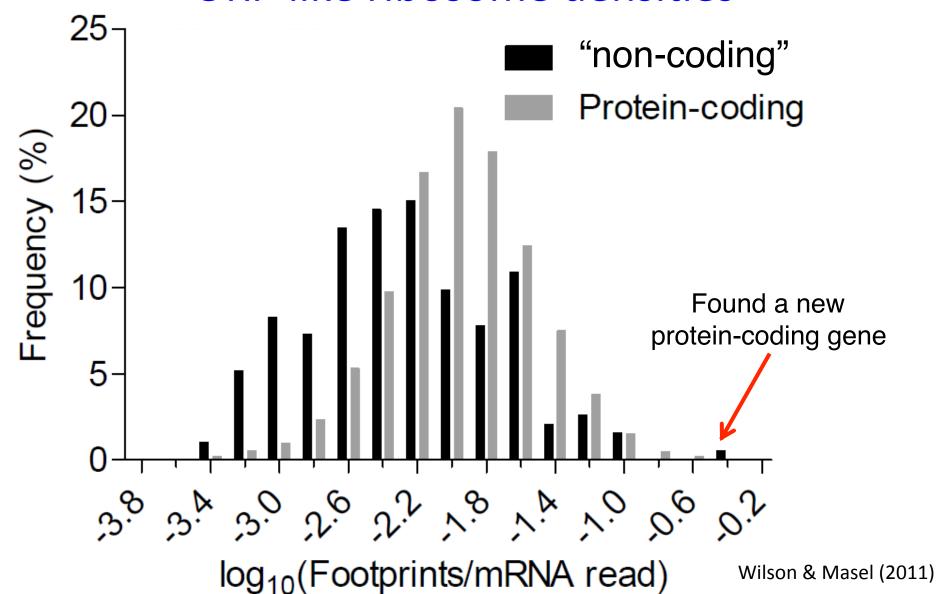


### Are "non-coding" transcripts associated with ribosomes?

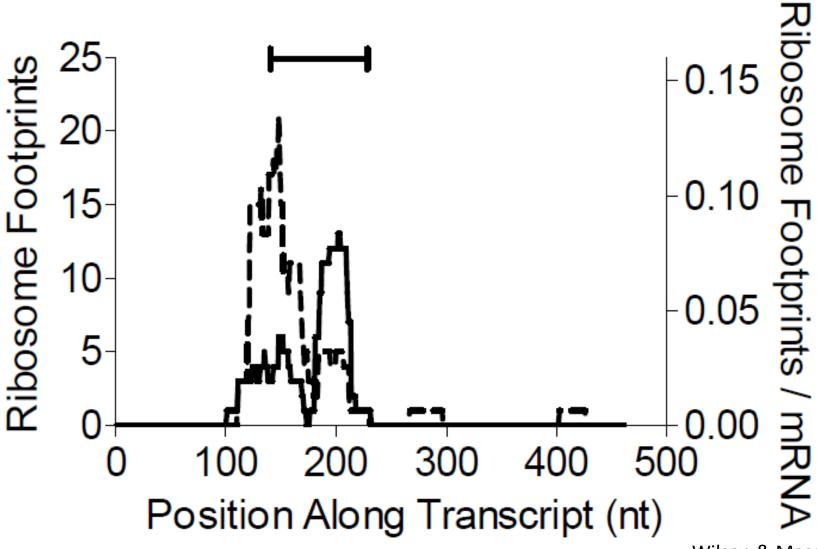
- Used ribosomal footprints that exactly mapped to unique genome site
   Ingolia et al. 2009
- 217/404 "non-coding" transcripts showed ribosomal association

Wilson & Masel 2011

### Many individual "non-coding" transcripts have ORF-like ribosome densities



## Ribosomal footprint locations match a 28aa ORF



#### Summary of ribosome profiling results

- Looks like a new coding sequence, but we don't know if polypeptide is functional
- Looks like de novo evolution
- Proof of principle of powerful method to annotate short de novo proteins
- Penultimate stage of gene birth is widespread

#### Conclusions

- Molecular errors are common and important (eg PPIs)
- 2 solutions to many molecular errors
  - low error rate via a proofreading mechanism for all sites
  - high error rate, but robustness to each separate error
- High error rates pre-screen future variants, and so promote evolvability
- With multiple loci, genetic diversity is not required for evolvability
- De novo genes may have been prescreened by widespread ribosomal association to "non-coding" sequences

### Broader picture

- Waste and mess and errors are not just a typical biological nuisance
- Without waste and mess, creative evolutionary innovations may not be possible
- Looking for a clean molecular machine can miss the essence of biology

### Thanks!

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John Templeton Foundation

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